



## Chemical Investigation and Antimicrobial Activity of *Francoeuria crispa* (Forssk) Grown Wild In Egypt

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- ✓ piperitone

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### Abstract

Survey and evaluation of wild plants that grow in Egyptian desert are of the priorities of the current study, so this present survey was carried out to examine the composition and antimicrobial activity of *Francoeuria crispa* (Forssk) plant growing wild in eastern Egyptian desert. *Francoeuria crispa* is an annual herb or sometimes a perennial subshrub producing small bright yellow flowers. The plant materials were collected from the plants growing on eastern desert approximately 1200 km out of Cairo. The chemical composition of the volatile oil, isolated by hydrodistillation of the herb, was investigated by GC and GC-MS. The percentage of essential oil of *F. crispa* (Forssk.), obtained in 0.23% v/w on a dry weight basis, consisted approximately of piperitone 45.0 %. Other oxygenated monoterpene compounds were found; the principal members were, cis-sabinene hydrate, (0.35%) linalool (0.85%)  $\alpha$ -terpineol (1.08%), neral (0.80%), linalyl acetate (0.47%) and, anethole trans (1.0%). Oxygenated monoterpene (OMG) group gave the highest percentage (50.85%) against 6.69, 4.8 and 27.77% for monoterpene hydrocarbon group (MHG), sesquiterpene hydrocarbon group (SHG) and oxygenated sesquiterpene group (OSG) respectively. The results proved that, *F. crispa* is rich in carbohydrates or glycoside, sterols and flavonoids. Tannins and coumarins are found in moderate amounts, while, alkaloids, saponins and anthraquinones are absent. The antimicrobial activities of the water and alcohol extracts of *F. crispa* herb were carried out using the inverted Petri plate method. The water and alcohol extraction showed prominent antimicrobial activities against fungi, gram positive and gram negative bacteria at a very low concentration (10  $\mu$ l).

## 1. Introduction

*Francoeuria crispa* (Forssk) syn. *Pulicaria crispa* (Forssk.), *Pulicaria undulata* (L.) (family Asteraceae) is an annual herb or sometimes a perennial subshrub producing small bright yellow flowers. This species is distributed in Egypt, Saudi Arabia, Kuwait, Iraq, Iran, Pakistan, India, Afghanistan and parts of North and West Africa [1-2]. *Francoeuria crispa* is an aromatic herb used in folk medicine in Egypt and Saudi Arabia for the treatment of inflammation. It is also repellent to insects [3] and it used as an herbal tea [4]. The constituents of *P. undulata* (L.) from Saudi Arabia was investigated by GC/MS. The *P. undulata* (L.) oil is rich in phenolic compounds and monoterpene hydrocarbons (MHG) with low incidence of sesquiterpenes hydrocarbons (SHG) [5]. The oil of *P. undulata* aerial parts exhibited inhibitory activity against Gram-positive and Gram-negative bacteria [6]. Undoubtedly, there is a clear and great interest in the moment of medicinal and aromatic plants as an important source of raw materials for the pharmaceutical treatment of many diseases and as an alternative to the use of chemicals. This is mainly due to the strong biological activity, where it is considered as safe, economical and powerful natural antioxidant [7]. Many investigators reported that, secondary metabolites are known to have many the therapeutic activities against many diseases in human categories; therefore, traditional medicinal plants can be used for treating many diseases [8-14] This present work was carried out to study the essential oil content, phytochemical screening and antimicrobial activities of *F. crispa* wild grown under Egyptian conditions.

## 2. Materials and methods

### 2.1. Plant material

The areal parts were collected from wild plant populations of *Francoeuria crispa* (Forssk) growing in sandy soils on Gebel Elba region approximately 1200 km south of Cairo in March 2013 (During flowering stage). Identification of the species was achieved by Prof. Dr. Loutfy Boulos [15-16]. Voucher specimens are in the herbarium of NRC, Cairo, Egypt.

## 2.2. Essential oil isolation

Five hundred grams from dried herb were subjected to hydro-distillation for 3 hrs using a Clevenger-type apparatus [17]. The essential oil content was calculated as a relative percentage (v/w). The samples of the essential oils were dehydrated over anhydrous sodium sulfate and stored in refrigerator until analyzed.

## 2.3. Gas chromatography (GC)

GC analyses were performed using a Shimadzu GC- 9A gas chromatograph equipped with a DB5 fused silica column (30 m x 0.25 mm i.d., film thickness 0.25 µm). Oven temperature was held at 40°C for 5 min and then programmed until 250°C at a rate of 4°C/min. Injector and detector (FID) temperature were 260°C; helium was used as carrier gas with a linear velocity of 32 cm/s.

## 2.4 Gas chromatography-Mass spectrometry (GC-MS)

GC-MS analyses were carried out on a Varian 3400 system equipped with a DB-5 fused silica column (30 m x 0.25 mm i.d.); Oven temperature was 40 to 240°C at a rate of 4°C/min, transfer line temperature 260°C, injector temperature 250°C, carrier gas helium with a linear velocity of 31.5 cm/s, split ratio 1/60, flow rate 1.1 ml/ min, Ionization energy 70 eV; scan time 1 mass range 40-350 amu.

## 2.5. Qualitative and quantitative analyses of essential oil

Identifications were made by library searches (Adams, 1995) [18] combining MS and retention data of authentic compounds by comparison of their GC retention indices (RI) with those of the literature or with those of standards available in our laboratories. The retention indices were determined in relation to a homologous series of n-alkanes (C8–C22) [19] under the same operating conditions. Further identification was made by comparison of their mass spectra on both columns with those stored in NIST 98 and Wiley5 Libraries or with mass spectra from literature. Component relative concentrations were calculated based on GC peak areas without using correction factors.

## 2.6. Preparation of the crude extracts

### 2.6.1. Alcoholic extract

The 80% ethanolic extracts are prepared following the process described [20]; 100 g of the aerial parts of each plant were collected, dried in the oven at 40 °C and reduced to powder. They were separately macerated with the 80% ethanol and allowed to stand for 72 hrs and then filtered. The filtrates were then evaporated under reduced pressure and dried using a rotary evaporator at 50°C. Dried extracts were stored in labeled sterile screw capped bottles at 5°C in the refrigerator, until when required for use.

### 2.6.2. Water extract

One hundred gm of the dried powder of each plant were macerated in water at room temperature for 24 hrs. The macerates were filtered and evaporated under vacuum till dryness. The residues were kept for testing and antimicrobial activity

## 2.7. Phytochemical screening:

The powdered air-dried aerial parts of plant under study was screened for carbohydrates and / or glycosides; sterols and / or triterpenes, flavonoids, tannins, saponins, coumarins and alkaloids, applying chemical tests [21]

## 2.8. Biological Activity:

### 2.8.1 Plant extraction

The dry aerial parts of the tested herb were extracted by 80% alcohol. The extract was evaporated under vacuum and the residue was dissolved in alcohol to give concentration of 100µg/ml.

### 2.8.2. Microbiological techniques

Microbial strains: The antimicrobial activity of alcoholic and water extracts were tested against some bacterial strains (Gram negative bacteria (G-): *Escherichia coli*, *Proteus vulgaris*, *Salmonella typhi*. (Gram positive bacteria (G+): *Bacillus subtilis*, *Pseudomonas fluroscens*, *Lactobacillus breveys*, *Staphylococcus aureus* and *chromobacter sp*), one fungal strain (*Aspergillus niger*) and one yeast strain (*Candida albicans*). Tested organisms were obtained from the Faculty of Agriculture, Cairo University.

## 2.9. Agar diffusion method

This method agar diffusion assay was carried out [22]. Nutrient agar was used for the cultivation of bacteria and yeast; while Czapek-Dox's medium (Dox 1910) was used for cultivation of fungal species. In this method, pre-sterilized Whatman no.1 filter paper discs (5 mm in diameter) (Whatman International Ltd., Maidstone, England) were impregnated with 100 µl of the extract (100 µg/ml) and was allowed to dry (to get rid of the alcohol) and was then applied on the surface of agar plates freshly seeded with standard inoculate of young

cultures, 24-hrs-old bacteria and yeast, and 7-days-old fungi. The plates of test organisms were then incubated at 27°C for 24 hrs for bacteria & yeast and for 48 hrs for fungi. At the end of the incubation period, the inhibition zones were measured (results are the average of triplicate measurements).

#### 2.10. Statistical analysis of data

All values of the antimicrobial activity were expressed as the mean of inhibition zones (mm) with three replicates for each treatment. Data were subjected to a paired-sample *t*-test using SPSS (ver. 9.0). *P*<0.05 was regarded as significant.

### 3. Results and discussion

#### 3.1. Preliminary phytochemical screening

Data presented in Table (1) show the preliminary phytochemical screening of the dried aerial parts of *F. crispa* in the flowering stage. Phytochemical examination of the alcoholic extracts showed that *F. crispa* is rich in volatile oils (natural compounds, characterized by, a strong aromatic odor), carbohydrates (energy source of living creatures), Sterols and/or terpenes (natural fatty substances) and flavonoids (antioxidant and anti-inflammatory substances). In addition, the dried aerial parts of *F. crispa* contain low amounts of tannins and coumarins. At the same time data in Table 1 confirm the absence of alkaloids, saponins and anthraquinones.

**Table 1.** Results of preliminary phytochemical screening of *Francoeuria crispa*

Chemical group	Phytochemical screening
Volatile oils	high value (++)
Carbohydrates and / or glycosides	high value (++)
Sterols and or terpenes	high value(++)
Flavonoids	high value(++)
Tannins	Low Value (+)
Alkaloids	Absent (-)
Saponins	Absent (-)
Coumarins	Low value (+)
Anthraquinones	Absent(-)

#### 3-2 Essential oil constituents of *F. crispa*

The essential oil extracted by water distillation from aerial parts of *F. crispa* yielded 0.23% (v/w) based on a dry weight. Twenty-three compounds representing 90.11% of the Egyptian *F. crispa* essential oil were identified (Table 2). The main constituents of Egyptian *F. crispa* oil are piperitone (45.0%), elemol (8.59%),  $\beta$ -eudesmol (8.17%),  $\alpha$ -eudesmol (5.35%), *cis*- nerolidol (4.5%),  $\alpha$ -terpinene (3.89%). Essential oil of *F. crispa* from Egypt grouped into four groups which are monoterpene hydrocarbons group (MHG), oxygenated monoterpene group (OMG), sesquiterpene hydrocarbons group (SHG) and oxygenated sesquiterpene group (OSG). The data in the same Table indicated that the (OMG) gave the highest concentrations (50.85%), followed by (OSG) (27.77%), (MHG) (0.69%) and (SHG) (4.8%). (OMG) included *cis*-sabinene hydrate, linalool,  $\alpha$ -terpineol, neral, geranal, piperitone, linalyl acetate and *trans*-anethole.  $\alpha$ -terpinene (3.89%) is the major constituent of the (MHG) of *F. crispa* essential oil. Other minor compounds in this group were detected as,  $\alpha$ -pinene, sabinene, 2-carene, limonene,  $\gamma$ -terpinene and 2,5-dimethyl styrene. The main constituents of (SHG) were *cis*- nerolidol, elemol, caryophyllene oxide,  $\alpha$ -eudesmol and  $\beta$ - eudesmol. These compounds amounted to 4.5, 8.59, 1.16, 5.35 and 8.17%, respectively. From the above data, it was observed that there are significant differences in the composition of *F. crispa* essential oil produced under Egyptian conditions in comparison with the oil produced under the other conditions [23], especially the contents of the major components. In Egypt the main constants of *F. crispa* were piperitone elemol and eudesmol, while, under the Iran conditions, the three major constituents were 4-terpineole (20.12%),  $\gamma$ -terpinene (7.0%) and  $\alpha$ -terpinene (4.02%). These results are consistent with the Szeveni-Galambosi *et. al.* (1992) [24] who proposed some chemo-types of some aromatic plants based on the dissimilar biosynthetic pathways which reflected the chemical contents of the essential oils formed under different conditions (different regions).Therefore, terpenoids as natural products differs constitutively by regions with different conditions.

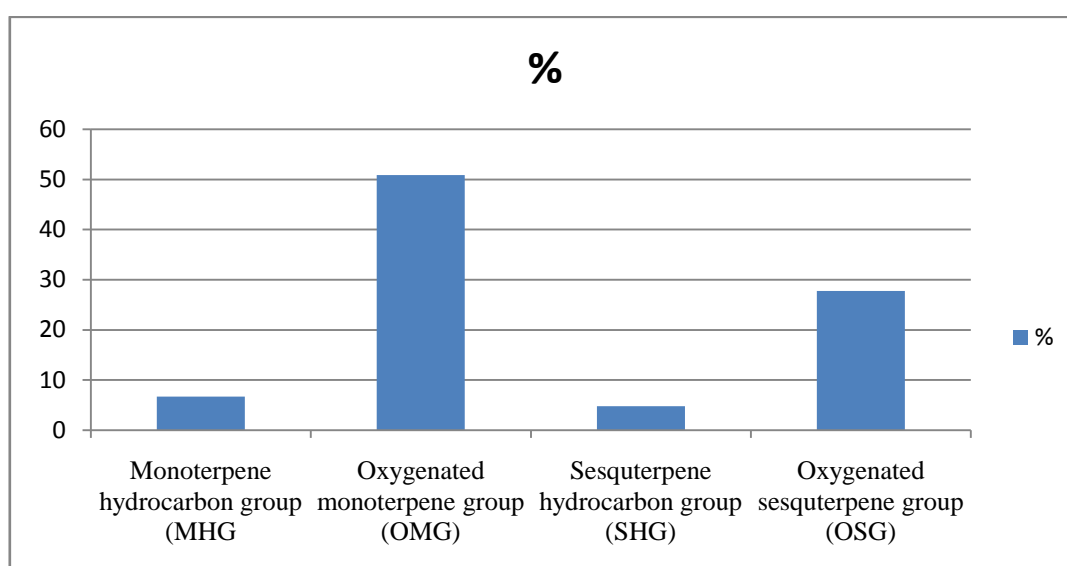
#### 2 Antimicrobial activities

The Antimicrobial activity was tested using gram negative bacteria: *Escherichia coli*, *Proteus vulgaris*, and *Pseudomonas fluorescens Salmonella typhi*, gram positive bacteria: *Bacillus subtilis*, *Chromobacter sp*, *Lactobacillus breveys*, *Staphylococcus aureus*, yeast: *Candida albicans* and fungi *Aspergillus niger*. Data in

Table 3 recorded that, the investigated water plant extract was effective against *E. coli* (inhibition zone 12.00 mm in diameter) against (15.00 mm) for 80 % alcoholic extract.

**Table 2.** Chemical constituents of essential oil extracted from *Francoeuria crispa* plant

Peak No	Compound	KI	%
<i>Monoterpene hydrocarbon group (MHG)</i>			
1	$\alpha$ -Pinene	939	0.23
2	Sabinene	976	0.17
3	2-Carene	1001	1.33
4	$\alpha$ -Terpinene	1018	3.89
5	Limonene	1039	0.31
6	$\gamma$ -Terpinene	1062	0.53
7	2,5 Dimethylstyrene	1096	0.23
Total (MHG)			6.69
<i>Oxygenated monoterpene group (OMG)</i>			
1	<i>cis</i> -Sabinene hydrate	1097	0.35
2	Linalool	1098	0.85
3	$\alpha$ -Terpineol	1189	1.08
4	Neral	1240	0.80
5	Geranal	1241	1.30
6	Piperitone	1252	45.0
7	Linalyl acetate	1257	0.473
8	<i>trans</i> -Anethole	1283	1.00
Total (OMG)			50.85
<i>Sesquiterpene hydrocarbon group (SHG)</i>			
1	$\beta$ -Caryophyllene	1418	0.75
2	$\beta$ -Gurjunene	1432	2.95
3	$\gamma$ -Muurolene	1477	1.10
Total (SHG)			4.80
<i>Oxygenated sesquiterpene group (OSG)</i>			
1	<i>cis</i> Nerolidole	1525	4.50
2	Elemol	1549	8.59
3	Caryophyllene oxide	1581	1.16
4	$\alpha$ -Eudesmol	1652	5.35
5	$\beta$ - Eudesmol	1652	8.17
Total(SOG)			27.77
Total contents			90.11



**Fig 1 :** Percentage of different chemical groups in *Francoeuria crispa* oil.

**Table3.** Antimicrobial assay of *F. crista* growing wild plant in eastern Egyptian desert.

Strains	Water extract	80 % alcoholic extract	Standard 100µg/disk
Gram-negative bacteria (G-)			
	Inhibition zone (mm in diameter) ± SE		
<i>Escherichia coli</i>	12.00 ± 0.65	15 ± 1.13	16.0 ± 0.6
<i>Proteus vulgaris</i>	0	0	21 ± 0.90
<i>Pseudomonas fluorescens</i>	0	0	26 ± 0.39
<i>Salmonella typhi</i>	0	0	19 ± 0.83
Gram -positive bacteria (G+)			
<i>Bacillus subtilis</i>	12.00 ± 0.5	14.00 ± 1.20	24 ± 0.51
<i>Chromobacter sp</i>	0	+	NT
<i>Lactobacillus breveis</i>	0	7.00 ± 0.33	NT
<i>Staphylococcus aureus</i>	0	0	22 ± 0.80
Yeast			
<i>Candida albicans</i>	0	0	12 ± 0.53
Fungi			
<i>Aspergillus niger</i>	12 ± 43	0	9.0 ± 0.01

0= not active + = mild effect NT= not tested; Values were expressed as mean ± SD; (Diameter on inhibition zone including well diameter of 6mm), standard for bacteria: amoxicillin, standard for fungi and yeast: canestin .

At the same time, the water extract was effective on *B. subtilis* (12.00 mm) while the alcoholic extract recorded (14.00 mm). Also there was an impact on the strain of *L. breveis* (7.00 mm) due to using 80 % alcoholic extract, while there was no effect observed for water extracts of the plant on the same strain, Obvious effect was observed with water extract of *F. crista* plant on the strain of fungi (*A. niger*) compared to non-existent effect of the alcoholic extract. Antimicrobial activity was previously investigated. The cited literature reported several biological benefits of wild plants [25-29]

## Conclusion

Dry herb of *F. crista* in the flowering stage has various chemical groups. It is rich in volatile oil, carbohydrates, Sterols and/or terpenes and flavonoids. It contains also low amounts of tannins and coumarins. At the same time, the data confirm the absence of alkaloids, saponins and anthraquinones. Twenty-three compounds representing 90.11% of the Egyptian *F. crista* essential oil were identified. The main constituents of piperitone (45.0%), elemol (8.59%), β-eudesmol (8.17%), α-eudesmol (5.35%), cis nerolidol (4.5%) and α-terpinene (3.89%). Other minor compounds were recorded. The antimicrobial activity was tested using gram negative bacteria, gram positive bacteria, yeast and fungi. Water extract of plants under investigation was effective on *E. coli* (12.00) and the inhibition zone was increased up to (15.00) for 80 % alcoholic extract. At the same time, the water extract was effective on *B. subtilis* (12.00) while the alcoholic extract recorded (14.00 mm). Also there was an effect on the strain of *L. breveis* (7.00 mm) due to using 80 % alcoholic extract, while there was no effect of water extracts of the plant on the same strain, Obvious effect was observed with water extract on the strain of fungi (*A. niger*) compared to non-existent effect of the alcoholic extract . In all cases, in traditional medicine, whole plants or plant mixtures are used instead of isolated compounds. There is evidence that crude plant extracts often have greater in vitro or/and in vivo antimicrobial activity than isolated constituents at an equivalent dose. However wild plants have many medicinal uses makes it very useful in the treatment of human diseases [30].

## References

1. L. Boulos, *Flora of Egypt* .Al-Hadara publishing. Cairo, Egypt. ISBN:???? 3 (2002)
2. A. Al-Rawi, *Flora of Kuwait*.. Alden press, Oxford. ISBN:???? 2 (1987).
3. M. Stavri, K.T. Mathew, A. Gordon, D. Steven, R. Shnyder, A. Falconer, S. Gibbons, *Oliv. Phytochem.*, 69 (2008) 1915
4. S.A. Ross, K.A El Sayed, M.A. El Sohly, M.T. Hamann, O.B. Abdel-Halim, A.F. Ahmed, M.M Ahmed *Planta Med.*, 63 (1997) 479
5. J.S. Mossa, M.S. Hifnawy, A.L. Yma, A.L. Mia, A.G. Mekkawi *Pharm. Biol.*, 25(1987) 113
6. H.H. EL-Kamali, A.H. Ahmed., A.S. Mohammed, A.M. Yahia., EL Tih, A.A. Ali, *Fitoterapia*. LXIX, (1998) 77
7. M.B. Tadhani, V.H. Petal, R.S. Subhash. *J. Food Compost. Anal.*, 20 (2007): 323.

8. Y. Li, C. Guo; J Yang, J Wei, J Xu, S Cheng, *Food Chem.*, 96 (2006) 254
9. FZ. Sabri, M.Belarbi, S. Sabri, MS. Alsayadi, *J. Nat. Prod. Plant Resour.*, 2 (2012) 512.
10. H. H. El-Kamali, and S. A.Mahjoub, *Ethnobotanical Leaflets* 13: 722-33. 2009
11. F.E. Hanbali, M.Akssira, A. Ezoubeiri, F., Mellouki, A.Benherraf, A.M. Blazquez, H. Boira, J Ethnopharmacol. 99 (2005) 399-401.
12. M. Znini, G. Cristofari, L. Majidi, J. Paolini, J. Desjobert, J.Costa, *LWT-Food Science and Technology*, 54 (2013) 564-569
13. N. Ali, F.S. Sharopov, M. Alhaj, G.M. Hill, A. Porzel, N. Arnold, W.N. Setzer, J.Schmidt, L. Wessjohann, *Nat Prod Commun.* 7 (2012) 257-260.
14. A. Javadinamin, J.Asgarpanah, *J. Essential Oil Bearing Plants*, 17(2014) 875-879.
15. L. Boulos, *Flora of Egypt: A Checklist*. Al-Hadara Publishing, Cairo. ISBN:???? (1995)
16. , L. Boulos, *Flora of Egypt VII Al Hadara Publishing Cairo, Egypt* ISBN:???? (2000)
17. J. F. Clevenger, *J Am Pharm Assoc* 17 (1928) 346.
18. R. P.Adams, *Allured, Carol Stream, Illinois, and U.S.A.* ISBN:???? (1995).
19. E. Kováts, *Gas-chromatographische charakterisierung organist verbindungen. Teil 1* ISBN:????
20. E. M. Williamson, D.T. Okpako, F.J.Evans, *John Wiley & Sons, Chichester*, ISBN:???? (1998) 15.
21. J. B. Harborne, *Phytochemical Methods by Chapman and Hall Ltd. London* ISBN:???? (1973).
22. CT. Collins, PM. Lyne, *Microbiological Methods (5th Edn), Butterworth and Co Pub Ltd, London and Toronto*, ISBN:???? (1985) 167
23. R. Mehdi, V.Jafar, N. Meissam, K. Mozghan, *J. Med. Plants Res.* 5 (2011) .
24. S. Galambosi, Z.; B. Galambosi; Y. Holm. *Finland.J. Essent. Oil Res.* 4 (1992) 375.
25. J. B. Harborne, *Phytochemical Methods by Chapman and Hall Ltd. London* (1973).
26. F. R. Anwar, U. Pak, *J. Bot.* 39 (2007)1443
27. S.S.Rani, S.P. Rao, M. Krishna, *JPRHC* 1(2009) 97.
28. H.Adbel baky, G. El baroty, *Inter. J. Econ. and Bus. Res.* 2 (2013) 99
29. H E. Yasmin, AM. Mona, *AJRC* 3(2015)58
30. R. Philippe, W. Colin, L W. Merlin, G. Ben, *Malar. J.* 10 (2011) 1.

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